Overview

This document explains how to stain polyacrylamide gels, comes directly from the Bio-Rad website, and can be used with catalog number: 161-0786.

Additional resources

Need more help?

Check the resources, and then see Ken

Main content

**SDS-PAGE Mini Gels**

1. Wash gel 3 times for 5 minutes, each in 200mL of ddH2O per gel.
2. Remove all of the water from the staining container and add 50mL Bio-safe Coomassie Stain (or enough to completely cover the gel)
3. Gently Shake for 1 hour.
   * Protein bands will be visible within 20 minutes and reach maximum intensity within 1 hour.
   * Longer incubations will not increase background.
4. Rinse gel in 200mL of ddH2O. for at least 30 minutes.
   * Stained gels can be stored in water
   * Residual SDS in the gel may cause background staining and interfere with band intensity while the gel is in the stain. Rinsing the gel extensively with water after staining will remove background and allow proper visualization of the bands.

**SDS-PAGE Gels**

1. For large gels (16-20”) increase volumes in above method 2 fold.

**Peptide Gels**

1. Fix gels in 40% methanol, 10% acetic acid for 30 minutes
2. Follow with the staining starting at step 2 above.
3. Extend step 4 water wash to two hours.
   * Peptide bands will not be clearly visible until after the final water wash